Integrated Workflow for Blood Collection, ccfDNA Stabilization and Extraction for **Non-Invasive Prenatal Testing** ¹Andrea Ullius, ¹Thorsten Voss, ²Stephan Busche, ²Wera Hofmann, ¹Daniel Grölz

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Background

Non-invasive prenatal testing (NIPT) uses fetal circulating cell-free DNA (ccfDNA) in the maternal blood circulation. The release of maternal genomic DNA from blood cells during blood collection, transport and storage significantly reduces the sensitivity of the next-generation sequencing (NGS) based NIPT assays. Moreover, the reliable and reproducible extraction of the low abundance and short fetal ccfDNA fragments is technically challenging. Both difficulties are addressed by the PAXgene® Blood ccfDNA System*, which consists of a blood collection tube (PAXgene Blood ccfDNA Tube*) and a ccfDNA purification kit (QIAsymphony® PAXgene Blood ccfDNA Kit*). The PAXgene Blood ccfDNA Tube uses a plastic BD Vacutainer[®] tube with a unique, non-crosslinking chemistry for stabilization of blood cells. The QIAsymphony PAXgene Blood ccfDNA Kit can be used for automated isolation of either small ccfDNA fragments or co-isolation of small and large ccfDNA fragments. The performance of the new system was evaluated in two research studies in comparison to K₂EDTA tubes and the Streck Cell-Free DNA BCT[®].



Figure 1. Standard and large fragment protocol lines. With the standard protocols (STA) predominately small ccfDNA fragments are isolated while large fragment protocols (LAF) efficiently co-purify large and small ccfDNA fragments. Both protocol lines consist of protocols for 2.4 ml or 4.8 ml plasma input.

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Study 1: Verification of the PAXgene Blood ccfDNA System

<u>Table 1</u>. System performance evaluation of the PAXgene Blood ccfDNA System **Relative ccfDNA Yield Relative ccfDNA Yield** Study 1A Manual Sample Preparation Automated Sample Preparation Comparison of PAXgene automated kit vs. manual procedure¹ **Study Parameters** Samples tested 825 y = 0.94x + 1.37v = 1.11x - 1.85Subjects $r R^2 = 0.91$ 189 R² = 0.91 Runs Instruments Study Results 104% ccfDNA yield Portion of dsDNA² > 96% **ຕົບ**^{- 10} Total failures Study 1B Evaluation of PAXgene system performance³ **Study Parameters** Samples tested 1636 20 Subjects 204 EDTA Tube C_T Values EDTA Tube C_T Values Runs Instruments **Figure 3.** Relative ccfDNA yield of paired plasma samples collected in PAXgene Blood ccfDNA Tubes as compared Study Results to EDTA tubes. Yield compared to EDTA plasma⁴ No significant difference C_{τ} values of a validated 18S rDNA PCR assay (precision range $\pm 3\sigma = 0.27 \Delta C_{\tau}$ indicated as dotted lines) are shown in ccfDNA stability at room temperature⁵ 7 days a scatter plot. ccfDNA was either isolated from plasma manually with the QIAGEN QIAamp Circulating Nucleic Acid **Cross-contamination** No evidence Kit (n = 60) (A) or with the QIAsymphony (QS) SP instrument using the QIAsymphony PAXgene Blood ccfDNA Kit Eluate freeze/thaw cycles and 4.8 ml plasma protocol for PAXgene Blood ccfDNA Tubes or the QIAGEN QIAsymphony Circulating DNA Kit for 192 Sample throughput per day⁶ EDTA tubes (n = 180) (B). 6 weeks Open reagent cartridge stability Runtime for 96 samples 5 h 40 min Instrument 2 **Instrument 3** Instrument ' ¹OIAamp Circulating Nucleic Acid Kit; ²Tested with restriction enzyme based assay for subset of eluates; OIAamp circulating Nucleic Acid Kit eluates with comparable portion of double-stranded DNA (dsDNA); ³Performance evaluated against RUO product specifications; ⁴Tested with 60 samples from 20 subjects, directly after blood collection; ⁵Tested with 60 samples from 20 subjects; ⁶Includes plasma processing and overnight runs **Shipment and Storage Simulation** Relative yield at day 3
Relative yield at day 7 2.5 -0



Figure 2. Relative ccfDNA yield compared to eluates generated from plasma separated directly after blood collection.

Plasma was generated from blood collected in PAXgene Blood ccfDNA Tubes after 5 hours of agitation (5 rpm) and 3 or 7 days of horizontal storage at 25°C to simulate typical transport and storage conditions. ccfDNA was purified manually using the QIAamp Circulating Nucleic Acid Kit.

Acceptance criteria: Relative yield must be \geq 0.5 and \leq 2.0 compared to Day 0, which equals, including the assay's imprecision [dotted line], a relative yield of \geq 0.41 and \leq 2.41.



Batch 1	neg.	22.11	neg.	22.44	neg.	23.45	Batch 1	neg.	22.44	neg.	22.45	neg.	22.65	Batch 1	neg.	22.27	neg.	22.28	neg.	22.28
7777	22.69	neg.	22.6	neg.	22.35	neg.	0.0000000	22.73	neg.	22.42	neg.	22.13	neg.		22.64	neg.	22.21	neg.	22.37	neg.
	neg.	22.14	neg.	22.66	neg.	23.27		neg.	22.38	neg.	22.38	neg.	22.48		neg.	22.36	neg.	22.34	neg.	22.25
	22.47	neg.	22.48	neg.	22.4	neg.		22.51	neg.	22.38	neg.	23.26	neg.		22.46	neg.	22.13	neg.	22.38	neg.
	777				-76												\mathbb{Z}/\mathbb{Z}			
Batch 2	neg.	neg.	neg.	neg.	neg.	neg.	Batch 2	neg.	neg.	neg.	neg.	neg.	neg.	Batch 2	neg.	neg.	neg.	neg.	neg.	neg.
7777	neg.	neg.	neg.	neg.	neg.	neg.		neg.	neg.	neg.	neg.	neg.	neg.		neg.	neg.	neg.	neg.	neg.	neg.
	neg.	neg.	neg.	neg.	neg.	neg.		neg.	neg.	neg.	neg.	neg.	neg.		neg.	neg.	neg.	neg.	neg.	neg.
	nea.	nea	nea	nea	nea	nea		nea	nea	nea	nea	nea	nea		nea	nea	nea	nea	nea	nea
		nog.	nog.	nog.	nog.	neg.		neg.	neg.	neg.	neg.	neg.	neg.		neg.	neg.	neg.	nog.	nog.	neg.
		nog.	nog.		1109.	nog.		neg.	neg.	neg.	neg.	neg.	neg.		neg.	nog.	neg.		nog.	neg.
Batch 3	22.31	neg.	22.35	neg.	22.42	neg.	Batch 3	22.61	neg.	22.35	neg.	22.34	neg.	Batch 3	22.38	neg.	22.29	neg.	22.21	neg.
Batch 3	22.31 neg.	neg. 22.16	22.35 neg.	neg.	22.42 neg.	neg. 22.14	Batch 3	22.61 neg.	neg. 22.04	22.35 neg.	neg. 22.42	22.34 neg.	neg. 22.33	Batch 3	22.38 neg.	neg. 22.25	22.29 neg.	neg. 22.21	22.21 neg.	neg.
Batch 3	22.31 neg. 22.37	neg. 22.16 neg.	22.35 neg. 22.34	neg. 22.17 neg.	22.42 neg. 22.08	neg. 22.14 neg.	Batch 3	22.61 neg. 23.33	neg. 22.04 neg.	22.35 neg. 22.58	neg. 22.42 neg.	22.34 neg. 22.51	neg. 22.33 neg.	Batch 3	22.38 neg. 22.48	neg. 22.25 neg.	22.29 neg. 22.32	neg. 22.21 neg.	22.21 neg. 22.37	neg. 22.34 neg.
Batch 3	22.31 neg. 22.37 neg.	neg. 22.16 neg. 22.15	22.35 neg. 22.34 neg.	neg. 22.17 neg. 30.18	22.42 neg. 22.08 neg.	neg. 22.14 neg. 22.14	Batch 3	22.61 neg. 23.33 neg.	neg. 22.04 neg. 22.24	22.35 neg. 22.58 neg.	neg. 22.42 neg. 22.54	neg. 22.34 neg. 22.51 neg.	neg. 22.33 neg. 22.27	Batch 3	22.38 neg. 22.48 neg.	neg. 22.25 neg. 22.23	neg. 22.29 neg. 22.32 neg.	neg. 22.21 neg. 22.23	22.21 neg. 22.37 neg.	neg. 22.34 neg. 22.36
Batch 3	22.31 neg. 22.37 neg.	neg. 22.16 neg. 22.15	22.35 neg. 22.34 neg.	neg. 22.17 neg. 30.18	22.42 neg. 22.08 neg.	neg. 22.14 neg. 22.14	Batch 3	22.61 neg. 23.33 neg.	neg. 22.04 neg. 22.24	22.35 neg. 22.58 neg.	neg. 22.42 neg. 22.54	22.34 neg. 22.51 neg.	neg. 22.33 neg. 22.27	Batch 3	22.38 neg. 22.48 neg.	neg. 22.25 neg. 22.23	22.29 neg. 22.32 neg.	neg. 22.21 neg. 22.23	22.21 neg. 22.37 neg.	neg. 22.34 neg. 22.36
Batch 3 Batch 4	22.31 neg. 22.37 neg. neg.	neg. 22.16 neg. 22.15 neg.	neg. 22.35 neg. 22.34 neg.	neg. 22.17 neg. 30.18 neg.	22.42 neg. 22.08 neg. neg.	neg. 22.14 neg. 22.14 neg.	Batch 3 Batch 4	22.61 neg. 23.33 neg.	neg. 22.04 neg. 22.24 neg.	neg. 22.35 neg. 22.58 neg.	neg. 22.42 neg. 22.54 neg.	neg. 22.34 neg. 22.51 neg.	neg. 22.33 neg. 22.27 neg.	Batch 3 Batch 4	22.38 neg. 22.48 neg. neg.	neg. 22.25 neg. 22.23 neg.	neg. 22.29 neg. 22.32 neg.	neg. 22.21 neg. 22.23 neg.	neg. 22.21 neg. 22.37 neg.	neg. 22.34 neg. 22.36 neg.
Batch 3 Batch 4	22.31 neg. 22.37 neg. neg. neg.	neg. 22.16 neg. 22.15 neg. neg.	neg. 22.35 neg. 22.34 neg. neg.	neg. 22.17 neg. 30.18 neg. neg.	22.42 neg. 22.08 neg. neg. neg.	neg. 22.14 neg. 22.14 neg. neg.	Batch 3 Batch 4	22.61 neg. 23.33 neg. neg.	neg. 22.04 neg. 22.24 neg. neg.	neg. 22.35 neg. 22.58 neg. neg.	neg. 22.42 neg. 22.54 neg. neg.	neg. 22.34 neg. 22.51 neg. neg.	neg. 22.33 neg. 22.27 neg. neg.	Batch 3 Batch 4	22.38 neg. 22.48 neg. neg. neg.	neg. 22.25 neg. 22.23 neg. neg.	neg. 22.29 neg. 22.32 neg. neg.	neg. 22.21 neg. 22.23 neg. neg.	neg. 22.21 neg. 22.37 neg. neg.	neg. 22.34 neg. 22.36 neg. neg.
Batch 3 Batch 4	22.31 neg. 22.37 neg. neg. neg. neg.	neg. 22.16 neg. 22.15 neg. neg. neg.	neg. 22.35 neg. 22.34 neg. neg. neg.	neg. 22.17 neg. 30.18 neg. neg.	22.42 neg. 22.08 neg. neg. neg.	neg. 22.14 neg. 22.14 neg. neg. neg.	Batch 3 Batch 4	22.61 neg. 23.33 neg. neg. neg. neg.	neg. 22.04 neg. 22.24 neg. neg. neg.	neg. 22.35 neg. 22.58 neg. neg. neg.	neg. 22.42 neg. 22.54 neg. neg. neg.	neg. 22.34 neg. 22.51 neg. neg. neg.	neg. 22.33 neg. 22.27 neg. neg. neg.	Batch 3 Batch 4	22.38 neg. 22.48 neg. neg. neg.	neg. 22.25 neg. 22.23 neg. neg. neg.	neg. 22.29 neg. 22.32 neg. neg. neg.	neg. 22.21 neg. 22.23 neg. neg. neg.	neg. 22.21 neg. 22.37 neg. neg. neg.	neg. 22.34 neg. 22.36 neg. neg. neg.

Figure 4: Cross Contamination Assessment.

C₋ values of a validated Y-chromosomal DYS14 PCR assay are shown. Three cross contamination test runs of 96 blood samples collected into PAXgene Blood ccfDNA Tubes, were carried out on 3 different QIAsymphony SP instruments using the QIAsymphony PAXgene kit and LAF protocol with 4.8 ml plasma. Per run 24 DYS14-positive (male) and 72 DYS14-negative (female) plasma samples were processed in a checkerboard setup interrupted by negative batches to test for batch to batch contamination.



Study 2: Validation of the PAXgene Blood ccfDNA System for non-invasive prenatal testing (NIPT) application

Study Setup:

- Blood samples were collected from 62 consented pregnant women
- Two blood collection tubes were collected per patient: Tube 1: Streck Cell-Free DNA BCT
- Tube 2: PAXgene Blood ccfDNA Tube
- Plasma processing from half of the blood samples collected into PAXgene tubes was performed with one centrifugation step, the remainder with two centrifugation steps
- All samples underwent the regular PrenaTest procedure per the standard assay protocol
- 25 plasma samples collected into PAXgene Blood ccfDNA Tubes were selected for downstream analysis (12 with one, 13 with two centrifugation steps)

QuantYfeX: Relative Fetal ccfDNA Content



Sample Number

Figure 5: Relative fetal fraction of paired plasma samples collected in PAXgene Blood ccfDNA Tubes or Streck **Cell-Free DNA BCT tubes.**

Relative amount of fetal ccfDNA in maternal background was quantified with methylation sensitive QuantYfeX QC Assay. Relative fetal ccfDNA fractions (PAXgene median: 13.7%; Streck BCT median: 12.4%) in maternal blood were not significantly different between the two different tubes (p-value: 0.20 paired T-Test, two-tailed distribution).



Sample Number



PAX Streck

Figure 6: NGS read out quality for paired plasma samples collected in PAXgene Blood ccfDNA Tubes or Streck **Cell-Free DNA BCT tubes.**

ccfDNA from Streck BCTs and PAXgene tubes sequenced on Illumina HiSeq[™] with NEB library preparation. There were significantly more mapped reads with ccfDNA from PAXgene tubes (p-value: 1.78E-04, two-tailed paired T-test; A).

There was no significant difference in unique mapped reads (B).

Study 2 Results

- Relative fetal ccfDNA fractions in maternal blood in both tubes were not significantly different
- 2. Significantly more mapped reads with ccfDNA from PAXgene tubes
- 3. No significant difference in uniquely mapped reads
- 4. No significant difference with PAXgene tube samples processed with one or two centrifugation steps
- 5. All samples which passed the internal quality standards produced equivalent results with the PrenaTest assay

Conclusions

Study 1:

- PAXgene Blood ccfDNA Tubes stabilize whole blood samples for up to 7 days of storage at room temperature (15–25°C) for efficient extraction of ccfDNA from plasma.
- Extraction of ccfDNA can be performed manually with the QIAamp Circulating Nucleic Acid Kit or automatically using the QIAsymphony PAXgene Blood ccfDNA Kit.
- Automated ccfDNA extraction with the QIAsymphony protocol offers minimal variation between replicates,
- highly reliable results and no cross-contamination.
- ccfDNA yield from PAXgene Blood ccfDNA Tubes stored for up to 7 days at room temperature is comparable to unstabilized EDTA blood on day 0.

Study 2:

- PAXgene Blood ccfDNA System fulfills requirements for whole blood stabilization and ccfDNA purification for NIPT.
- PAXgene Blood ccfDNA System is fully compatible with the NGS-based LifeCodexx PrenaTest.