



Circulating Cell-Free DNA Pre-analytics: The Importance of Standardized Workflows for Liquid Biopsy Applications

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Deficiencies in Routine Healthcare Call for Workflow Standardization



Diagnostic errors cause about 10% of all patient deaths and about 17% of adverse events

Institute of Medicine (IOM) Report Sept. 2015

The pre-analytical phase accounts for 46% to 68% of such errors observed during the total testing process

Medical Laboratory Observer, May 2014

SPIDIA & SPIDIA4P: Standardization and Improvement of Pre-analytical Procedures for *in vitro* Diagnostics *EU* FP7-HEALTH (GA. no. 222916) & *EU* HORIZON 2020 (GA. no. 733112)



SPIDIA4P

- QIAGEN coordinating both initiatives (16 & 19 Partners)
- 22 CEN Technical Specifications and ISO Standards planned or developed (highly consensus-driven international and European processes)
- E.g., Specifications for pre-examination processes for venous whole blood in Molecular in vitro diagnostic examinations
 - Part 3: Isolated circulating cell-free DNA from plasma CEN/TS 16835-3: 2015

Sample to Insight



Critical Points Along the Workflow







ccfDNA was extracted from EDTA plasma of 1 subject directly after blood draw (t0). 1 µl eluate was analyzed using the Agilent[®] High Sensitivity DNA Kit.





ccfDNA was extracted from EDTA plasma of 1 subject directly after blood draw (t0) and after 6 days at room temperature (t6). 1 µl eluate was analyzed using the Agilent[®] High Sensitivity DNA Kit.



PAXgene[®] Blood ccfDNA Tubes (RUO)* help prevent release of gDNA into plasma



ccfDNA was extracted from EDTA and PAXgene plasma of 1 subject directly after blood draw (t0) and after 6 days at room temperature (t6). 1 µl eluate was analyzed using the Agilent[®] High Sensitivity DNA Kit.

 * For Research Use Only. Not for use in diagnostic procedures.



Unique stabilization of extracellular levels of ccfDNA

- Effective stabilization at RT minimizes background gDNA and maximizes ccfDNA yield from plasma
 - White blood cells helps prevent release of gDNA
 - Red blood cells helps minimize hemolysis
- Non-crosslinking NA preservation no DNA modification

BD Vacutainer[®] plastic tube with BD Hemogard[™] safety closure

- Helps minimize risk of tube breakage
- Enhanced safety for healthcare and lab personnel
- Helps minimize contamination between samples
- Provides consistent blood draw volume

Integrated pre-analytical workflow

 Seamless integration into manual or automated prep with QIAamp[®] and QIAsymphony[®] circulating DNA extraction technology



Sample to Insight



System Approach: QIAsymphony PAXgene Blood ccfDNA Kit (RUO)*



- Dedicated isolation technology for use with PAXgene Blood ccfDNA Tube (RUO)* — Complete system: ccfDNA stabilization + extraction
 - Binding chemistry optimized for use with PAXgene ccfDNA Tube reagent
 - Optimized input volumes to accomodate higher volume plasma
 - Optional custom protocols for primary tube handling
- Two protocol lines
 - Standard protocol similar to QIAsymphony DSP Circulating DNA Kit[†] protocols (≤500 bp)
 - Large fragment protocols enable co-isolation of large fragments (>500 bp) with flexible elution volume (60, 100, 150 μL)

⁺ Intended for in vitro diagnostic use.

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Validation for noninvasive prenatal testing (NIPT)



Equivalence between PAXgene Blood ccfDNA Tubes (RUO)* and current method



QuantYfeX[™]: Relative Fetal ccfDNA Content

Relative fetal fraction of paired plasma samples collected in PAXgene Blood ccfDNA Tubes or Streck Cell-Free DNA BCT tubes. Relative amount of fetal ccfDNA in maternal background was quantified with methylation sensitive QuantYfeX QC Assay. Relative fetal ccfDNA fractions in maternal blood were not significantly different between the two different tubes (p-value: 0.20 paired T-Test, two-tailed distribution). PAXgene median: 13.7% (samples 1-30 with 2 centr. 12.0%; samples 31-62 with 1 centr. 15.4%), Streck BCT: median 12.4% (median indicated as dotted lines). Data courtesy of LifeCodexx.

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Equivalence between PAXgene Blood ccfDNA Tubes (RUO)* and current method



NGS read out quality for paired plasma samples collected in PAXgene Blood ccfDNA Tubes or Streck Cell-Free DNA BCT

tubes. ccfDNA from Streck BCTs and PAXgene tubes sequenced on Illumina HiSeq[™] with NEB library preparation. There were significantly more mapped reads with ccfDNA from PAXgene tubes (p-value 1.78E-04, two-tailed paired T-test). PAXgene median: 90.95% (samples 1-13 with 2 centr. 90.95%; samples 14-25 with 1 centr. 90.92%), Streck BCT: median 89.37% (A). There was no significant difference in unique mapped reads. PAXgene median: 83.27% (samples 1-13 with 2 centr. 83.30%; samples 14-25 with 1 centr. 83.25%), Streck BCT: median 83.25% (B);

median indicated as dotted lines. Data courtesy of LifeCodexx.

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[†] For molecular biology applications. Not intended for the diagnosis, prevention or treatment of a disease.



ccfDNA from Plasma — Sample-to-Insight Workflows



Complete preanalytical workflow solutions from



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- [‡] Intended for in vitro diagnostic use.

Sample to Insight



Thank You!

